



# YSI 6131 and 6132 Blue-Green Algae Sensors

### Phycocyanin and Phycoerythrin Sensors

Blue-green algae (a. k. a. cyanobacteria) monitoring is of growing intrest due to the problems some species can present through the production of toxins and compounds that deteriorate the quality of drinking water and through the formation of blooms. Blue-green algae are of interest for ecosystem studies and monitoring as well, where

they may represent the most abundant primary producer.

• Two versions: One for freshwater (phycocyanin) and one for marine (phycoerythrin) environments

• Optimized for excellent sensitivity for monitoring algal populations at natural levels, providing an early warning for bloom conditions

• Insensitive to potential interferences including chlorophyll, turbidity, and dissolved organics

- Fully compatible with all YSI 6-Series sondes equipped with optical ports—with a free and easy firmware upgrade from ysi. com
- Integrated wiping system providing anti-fouling in the most hostile environments
- Durable mechanical features including a non-corroding titanium wiper shaft, service-center replaceable wiper shaft seal, and a new switch controlled wiper parking system to prevent mis-parking

The YSI 6131 Phycocyanin Blue-green Algae Sensor

## Phycocyanin Blue-Green Algae Sensor (BGA-PC)

The BGA-PC sensor should be used in freshwater and estuarine conditions. Common applications in these environments include:

- Monitoring taste and odor causing species in drinking water reservoirs
- Used as a possible presence of harmful or toxic species as part of HAB monitoring systems
- Ecosystem monitoring where blue-green algae are present

## Phycoerythrin Blue-Green Algae Sensor (BGA-PE)

The BGA-PE sensor should be used in marine and estuarine conditions. Common applications in these environments include:

- Ecosystem monitoring where blue-green algae may represent important primary producers.
- As a key sensor in a Harmful Algae Bloom monitoring system
- Nutrient cycling studies where blue-green algae are present





To order, or for more information, contact YSI Environmental.

+1 937 767 7241 800 897 4151 (US) www.ysi.com

YSI Environmental +1 937 767 7241 Fax +1 937 767 9353 environmental@ysi.com

Endeco/YSI +1 508 748 0366 Fax +1 508 748 2543 systems@ysi.com

SonTek/YSI +1 858 546 8327 Fax +1 858 546 8150 inquiry@sontek.com

YSI Gulf Coast +1 225 753 2650 Fax +1 225 753 8669 environmental@ysi.com

YSI Hydrodata (UK) +44 1462 673 581 Fax +44 1462 673 582 europe@ysi.com

YSI Middle East (Bahrain) +973 1753 6222 Fax +973 1753 6333 halsalem@ysi.com

YSI (Hong Kong) Limited +852 2891 8154 Fax +852 2834 0034 hongkong@ysi.com

YSI (China) Limited +86 10 5203 9675 Fax +86 10 5203 9679 beijing@ysi-china.com

YSI Nanotech (Japan) +81 44 222 0009 Fax +81 44 221 1102 nanotech@ysi.com

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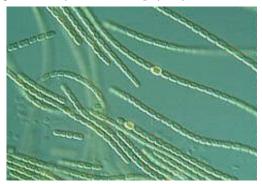
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### **Methodology**

The simple technique of in-vivo fluorometry (IVF) for locating and measuring algae in real-time has been in use by oceanographers and limnologists for over 30 years (Lorenzen, C.J., 1966) and is based on the direct measurement of the fluorescence of the chlorophyll a in the living algal cells. The same methodology is used to detect the phycobilin pigments found in blue-green algae (a.k.a. cyanobacteria), phycocyanin and

phycoerythrin. Blue-green algae pigment fluorescence is the only technique available that is sensitive enough to detect blue-green algae at natural levels without concentration or extraction. Because there is no special sample handling or processing required, IVF is ideal for profiling, moored and real-time data collection using YSI multiparameter sondes. Similar to the YSI chlorophyll sensor, the blue-green algae sensors do not provide quantitative pigment concentration data, but rather supply relative data on the biomass of blue-green algae. This data is extremely useful for tracking temporal or spatial changes in the distribution of the bluegreen algae population and to provide an early warning of increasing biomass that can lead to a bloom. IVF data can be correlated to quantitative data in order to calibrate



Anabaena sp., a common blue-green algae found in freshwater environments.

the IVF data to provide concentration estimates. Without correlation, IVF provides a relative cyanobacteria measure that can be used to track trends and trigger more specific tests.

Both Blue-Green Algae Sensors (BGA-PC and BGA-PE) have been optimized to be as sensitive as possible and used for monitoring natural populations of blue-green algae prior to a full bloom condition. The sensors should be used to monitor natural temporal or spatial fluctuations in BGA biomass as well as indicate the possible onset of an algal bloom. During an algal bloom, it is possible that the sensor will be over-range and cell concentrations could be in the millions of cells/mL.

## **Calibration Considerations**

True calibrations with a primary standard are not practical for IVF applications. IVF measures the relative change in cyanobacterial biomass via pigment fluorescence and the best means of calibration is to compare the IVF data with quantitative data generated from a water sample that was taken in the field when the IVF sensor was deployed. However, users should use a secondary standard to ensure that the sensor is stable and working properly before deployment.

Lorenzen, C. J., 1966. A Method for the Continuous Monitoring of In-Vivo Chlorophyll Concentration. Deep Sea Research, 13, 223-227.

		Range	Detection Limit	Resolution	Linearity
	BGA - Phycocyanin*	~0 to 280,000 cells/mL <sup>†</sup> 0 to 100 RFU	~220 cells/mL $^{\circ}$	1 cell/mL 0.1 RFU	R <sup>2</sup> > 0.9999*
	BGA - Phycoerythrin*	~0 to 200,000 cells/mL $^{\dagger}$ 0 to 100 RFU	~450 cells/mL <sup>§§</sup>	1 cell/mL 0.1 RFU	R <sup>2</sup> > 0.9999**
ed	• Maximum depth rating for all standard optical probes is 200 feet, 61 m. Also available in a Deep Depth option (0 to 200 m). BGA = Blue-Green Algae RFU = Relative Fluoresence Units ~ = Approximately	† Explanation of Ranges can be found in the 'Principles of Operation' section of the 6-Series Manual, Rev D.	\$ Estimated from culture \$\$ Estimated from culture	s of Microcystis aeruginosa. es Synechococcus sp.	*Relative to serial dilution of Rhodamine WT (0-400 ug/L). **Relative to serial dilution of Rhodamine WT (0-8 µg/L).